

# Prevention of Alcohol-Heightened Aggression by CRF-R1 Antagonists in Mice: Critical Role for DRN-PFC Serotonin Pathway

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Alcohol can escalate aggressive behavior in a significant subgroup of rodents, humans, and nonhuman primates. The present study investigated whether blockade of corticotropin-releasing factor receptor type I (CRF-R1) could prevent the emergence of alcohol-heightened aggression in mice. The serotonin (5-HT) pathway from the dorsal raphe nucleus (DRN) to the medial prefrontal cortex (mPFC) by CRF-R1 was investigated as a possible target for the prevention of alcohol-heightened aggressive behavior. Male CFW mice that reliably exhibited aggressive behaviors after consuming 1 g/kg of alcohol received systemic or intra-DRN administration of CRF-R1 antagonists, CP-154,526 or MTIP, before a confrontation with a male conspecific. Blockade of DRN CRF-R1 receptors with both antagonists significantly reduced only alcohol-heightened aggression, whereas systemic administration reduced both alcohol-heightened and species-typical aggression. Next, a 5-HT1A agonist, 8-OH-DPAT, was coadministered with CP-154,526 into the DRN to temporarily disrupt 5-HT activity. This manipulation abolished the antiaggressive effects of intra-DRN CP-154,526. In the mPFC, *in vivo* microdialysis revealed that extracellular 5-HT levels were increased in mice that consumed alcohol and were then injected with CP-154,526, both systemically or intra-DRN. Neither alcohol nor CP-154,526 alone affected 5-HT release in the mPFC. The present results suggest the DRN as a critical site for CRF-R1 to modulate alcohol-heightened aggression via action on the serotonergic DRN–PFC pathway.

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## INTRODUCTION

Intense outbursts of aggressive behavior have been linked to alcohol drinking in humans and other animal species (see, eg, Bushman and Cooper, 1990; Krug *et al*, 2002; Roizen, 1997). Evidence for alcohol as a promoter of escalated levels of aggressive behavior has been established in laboratory settings, particularly in susceptible subgroups of individuals (Cloninger, 1987; Miczek *et al*, 2004). In mice, low to moderate doses of alcohol can escalate aggressive behaviors to levels that are significantly higher and more injurious than species-typical aggression (Miczek *et al*, 1998; Miczek and de Almeida, 2001), modeling critical features of alcohol-escalated aggression.

Understanding the neurobiological mechanisms underlying escalated aggression, particularly those mediating alcohol-heightened aggression, has been a challenge. Con-

siderable evidence supports the involvement of serotonergic mechanisms in aggression; however, it remains controversial whether it is an *increase* or *decrease* of serotonin (5-HT) activity that promotes escalated aggression (Takahashi *et al*, 2011; de Boer and Koolhaas, 2005). Ascending serotonergic projections arise mainly from the dorsal raphe nucleus (DRN) to innervate forebrain and limbic regions (Jacobs and Azmitia, 1992; Molliver, 1987). Of particular interest, 5-HT levels in the medial prefrontal cortex (mPFC) are dynamically regulated in preparation for, during, and in recovery from an aggressive confrontation in rats (van Erp and Miczek, 2000). Phasic changes in 5-HT activity may also affect alcohol-heightened aggression and other forms of escalated aggression, as evidenced by infusion of drugs targeting 5-HT1 receptors into the DRN and subregions of the PFC (Bannai *et al*, 2007; de Almeida *et al*, 2006; Faccidomo *et al*, 2008).

DRN 5-HT neurons are finely tuned by different inputs, one of which is corticotropin-releasing factor (CRF; Valentino and Commons, 2005). CRF has two receptor subtypes, CRF-R1 and CRF-R2, both found in high densities in the DRN (Chalmers *et al*, 1995; Swanson *et al*, 1983). Infusion of CRF or drugs targeting CRF receptors in the DRN significantly affects the firing rate of 5-HT neurons in a dose-dependent manner (Kirby *et al*, 2000; Lowry *et al*,

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2000), and also controls 5-HT release in the PFC, striatum, and other limbic regions (Forster *et al*, 2008; Lukkes *et al*, 2008; Price and Lucki, 2001).

In the present study, we targeted CRF-R1 within the DRN and the 5-HT projections to the mPFC as a putative mechanism for the emergence of alcohol-heightened aggression. CRF mechanisms significantly modulate maternal and intermale aggression in mice, hamsters, and rats (Elkabir *et al*, 1990; Farrokhi *et al*, 2004; Gammie *et al*, 2004; Gammie and Stevenson, 2006; Tazi *et al*, 1987). Although evidence points to a key role of CRF-R1 in several features of alcohol-related behaviors and effects (Heilig and Koob, 2007), its involvement in alcohol-heightened aggression needs to be examined.

## MATERIALS AND METHODS

### Housing

Male CFW Swiss-derived mice (Charles River Laboratories, Wilmington, MA), ~6 weeks old at arrival, were housed in pairs with a female conspecific in standard polycarbonate cages (28 × 17 × 14 cm) for 3 weeks before any manipulation. Additional CFW male mice were housed in groups of 8–11 per cage (48 × 26 × 14 cm) and used as intruders described below. Animals were maintained in a temperature- and humidity-controlled room (21 ± 1 °C and 35–40%, respectively), with lights on from 1900 h to 0700 h. Food was available *ad libitum*, and access to water was limited to 4 h/day to enhance alcohol self-administration, as described below (Faccidomo *et al*, 2008). Experimental procedures were approved by the Institutional Animal Care and Use Committee of Tufts University, and followed the Guide for the Care and Use of Laboratory Animals (National Research Council of The National Academies, 2010).

### Drugs

Two nonpeptidergic, high-affinity, and selective CRF-R1 antagonists were used in these experiments. CP-154,526 was the first nonpeptidergic CRF-R1 antagonist described to effectively cross the blood–brain barrier (Schulz *et al*, 1996), with well-characterized pharmacological and behavioral effects in the literature (see Seymour *et al*, 2003). Our previous studies (Hwa *et al*, 2013) revealed that it was suitable for intra-DRN infusions as well as for systemic administration (intraperitoneal (i.p.)), enabling appropriate drug effect comparisons (systemic *vs* intracranial). MTIP is a more recently developed molecule, showing high affinity and specificity to CRF-R1 with no relevant affinity to any of 74 other receptor and channels tested, good oral bioavailability, important behavioral effects (similar to those of other CRF-R1 antagonists), and high promise for entering clinical trials at the time these experiments were carried out (Gehlert *et al*, 2007).

CP-154,526 (butyl-ethyl-[2,5-dimethyl-7-(2,4,6-trimethylphenyl)-7H-pyrrolo[2,3-d]pyrimidin-4-yl]-amine) was either donated (Pfizer, New York, NY) or commercially purchased (Tocris Bioscience, Bristol, UK). MTIP (3-(4-chloro-2-morpholin-4-yl-thiazol-5-yl)-8-(1-ethylpropyl)-2,6-dimethylimidazo[1,2-*b*]pyridazine) was kindly donated by Dr Markus Heilig (NIAAA/NIH). 8-OH-DPAT ((±)-8-hydroxy-2-(di-*n*-propylamino)tetralin) hydrobromide was purchased from

Sigma-Aldrich. For i.p. administration, CP-154,526 was dissolved in 0.1% carboxymethyl cellulose (CMC), and MTIP was dissolved in 2% Tween-80. For intracerebral administrations, CP-154,526, MTIP, and 8-OH-DPAT were dissolved in artificial CSF (aCSF).

### Alcohol Self-Administration

Experimental males were trained for alcohol self-administration using an operant conditioning panel inserted into their home cage (Miczek and de Almeida, 2001). The panel had two troughs with one connected to a syringe located behind the panel and filled with 6% alcohol or water. The infrared beam inside the troughs detected nose pokes, with every fifth nose poke (fixed ratio 5 (FR5)) delivering a 50- $\mu$ l droplet of either 6% alcohol solution (w/v) or water, as controlled by computer software (Med Associates, St Albans, VT). Self-administration was terminated when animals achieved the moderate dose of 1 g/kg of alcohol or equivalent volume of water (corresponding to 8–12 reinforcements, depending on body weight). Alcohol self-administration sessions were limited to twice a week to avoid extensive exposure to alcohol. Within 2–3 weeks of training, all mice completed alcohol or water reinforcement sessions within 4 min.

### Assessment of Aggression

Concomitant with self-administration training, experimental males established aggressive behavior against a naive group-housed intruder male (Miczek and O'Donnell, 1978). Briefly, each experimental male confronted an intruder in the experimental male's home cage, for 5 min after the first attack bite. The female was removed from the experimental male's home cage during confrontations. Frequency of attack bites was recorded to assess aggression levels. The criterion used for stable aggression levels was defined as the frequency of attack bites across 3 consecutive sessions varied within 15% for each experimental male (criterion met within 7–12 confrontations). Approximately 5% of mice were excluded from the study because of a failure in establishing stable and reliable aggressive behaviors.

### Alcohol-Heightened Aggression

After experimental males displayed stable levels of drinking and fighting, they were tested for aggression 15 min following 1 g/kg alcohol drinking (Miczek and de Almeida, 2001). For comparison purposes, baseline aggression levels were determined after consuming water. Confrontations were recorded and analyzed by a trained observer using The Observer software (Observer XT 8.0, Noldus Information Technology, Wageningen, The Netherlands). Frequency and duration of aggressive behaviors, such as attack bites and sideways threats, as well as nonaggressive composite behaviors, such as walking, rearing, and grooming, were determined (Miczek and O'Donnell, 1978). Aggressive confrontations were conducted in 48–72 h intervals, for a total of 6–8 aggression tests, alternating between water and alcohol tests. After completing these tests, experimental mice were subjected to the following drug experiments.

## Systemic Injections of CRF-R1 Antagonists

In two separate sets of mice, the effects of CRF-R1 receptor antagonists, CP-154,526 or MTIP, were assessed in aggressive mice after consuming water or 1 g/kg alcohol. On a test day, experimental mice completed self-administration of alcohol or water, and were immediately injected i.p. with CP-154,526 (vehicle, 10, 17, or 30 mg/kg,  $n = 18$ ), or MTIP (3, 10, or 30 mg/kg,  $n = 13$ ). At 15 min after drug treatment, mice confronted a male intruder, as described above. A within-group design was used, in which each mouse was examined under all treatment conditions, in a counter-balanced treatment order. Vehicle tests (2–3 tests) were interspersed between drug tests to ensure the absence of any carryover effects.

## Intra-DRN Microinjection of CRF-R1 Antagonists and 8-OH-DPAT

Separate subsets of experimental mice underwent intracerebral cannulation surgery for the intra-DRN microinjection studies (Takahashi *et al*, 2010a; Hwa *et al*, 2013). The mice were anesthetized with a mixture of ketamine (100 mg/kg) and xylazine (10 mg/kg), placed in a stereotaxic frame (David Kopf Instruments, Tujunga, CA), and implanted with 26-gauge guide cannulae (PlasticsOne, Roanoke, VA) aimed 2 mm above the DRN, AP:  $-4.2$ ; ML:  $\pm 1.5$ ; DV:  $-1.9$  mm relative to bregma, angled  $26^\circ$  to the vertical (Paxinos and Franklin, 2001). A 33-gauge obturator (PlasticsOne) was inserted into the guide cannula to prevent blockage or scarring, and was moved daily to habituate the mice to the handling procedure. After 5 days of recovery, experimental mice were reassessed for alcohol self-administration and aggression at least twice before microinjection experiments.

The procedures used in the study were based on previous work (Faccidomo *et al*, 2008; Takahashi *et al*, 2010a). On testing days, immediately after completing the drinking requirement, the obturator was removed and replaced with a 33-gauge microinjector (PlasticsOne) attached via polyethylene tubing (Intramedic PE-50) to a CMA/100 microinjection pump (CMA Microdialysis AB, Holliston, MA). The microinjector extended 2 mm beyond the tip of the guide cannula, through which vehicle (aCSF), CP-154,526 (0.3 or 0.6  $\mu$ g), or MTIP (0.345  $\mu$ g, matched molarity of 0.3  $\mu$ g CP-154,526 solution) were infused into the DRN (0.25  $\mu$ l over 2 min). Each drug (CP-154,526 or MTIP) was tested in separate cohorts of mice. Doses for microinjection were chosen based on a pilot study detecting promising effects after a 0.3  $\mu$ g infusion of CP-154,526 into the DRN, and parallel studies also showed relevant effects of microinjecting 0.3 and 0.6  $\mu$ g CP-154,526 on alcohol consumption, as recently published by Hwa *et al* (2013). The microinjector was left in place for an additional 1 min to allow the drug to diffuse. During the infusion, mice were left unrestrained. After 10 min, a male intruder was introduced into the experimental male's home cage, and a confrontation took place. Each experimental subject received a total of 6–8 microinjections, in counterbalanced order.

In a separate group of mice, 0.3  $\mu$ g 8-OH-DPAT was coadministered with CP-154,526. We used an autoreceptor agonist, 8-OH-DPAT, as a pharmacological tool to transi-

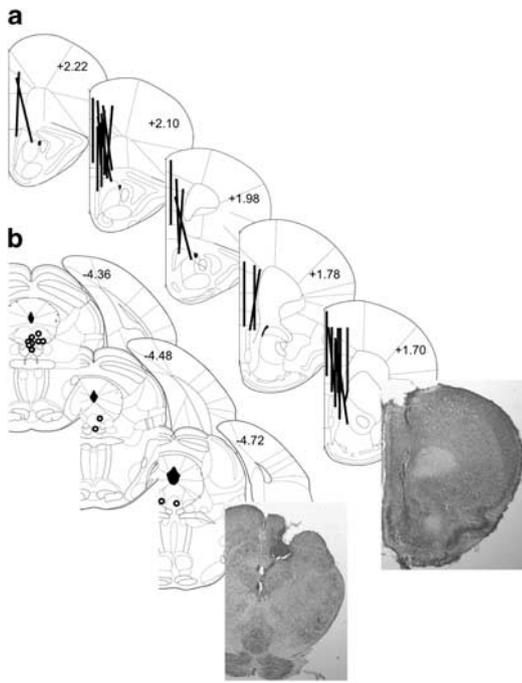
ently inhibit 5-HT impulse flow from the DRN (Sprouse and Aghajanian, 1987; Will *et al*, 2004) during testing for aggressive behavior. As intra-DRN infusion of 8-OH-DPAT has been reported to reduce aggressive and motor behaviors in mice in higher doses (1  $\mu$ g; Faccidomo *et al*, 2008), we selected a lower dose, 0.3  $\mu$ g, so that baseline aggression and motor activity would be preserved, as shown by Takahashi *et al* (2010b). After drinking water or 1.0 g/kg of alcohol, mice received intra-DRN microinfusions of 0.3  $\mu$ g CP-154,526 alone, 0.3  $\mu$ g 8-OH-DPAT alone, a combination of CP-154,526 and 8-OH-DPAT, or vehicle in counterbalanced order and subsequently tests for aggressive behavior commenced.

## 5-HT Concentrations in the mPFC

A separate cohort of mice that were trained for alcohol self-administration and for aggressive behaviors were implanted with a microdialysis CMA/7 guide cannula for 5-HT measurements in the mPFC (CMA Microdialysis AB) 1 mm above the mPFC, AP:  $+2.6$ ; ML:  $-0.3$ ; DV:  $-0.8$  mm to bregma. For the intra-DRN microinfusion experiment, animals were also implanted with an additional 26-gauge microinjection guide cannula aimed 2 mm above the DRN, as described above. Mice were allowed to recover from surgery for 1 week, during which they were handled daily. After alcohol self-administration was reestablished, a CMA/7 probe with a 1-mm active membrane was inserted into the mPFC under isoflurane inhalation anesthesia, and the probe was perfused overnight with aCSF at a flow rate of 0.5  $\mu$ l/min. On the following morning, the flow rate was increased to 1.5  $\mu$ l/min, and 20-min microdialysate samples were collected after 1 h stabilization. Mice were housed in their home cages during sample collection with no access to water or food to prevent further fluctuations in 5-HT release. Seven baseline samples were collected, and then the operant conditioning panel was inserted into the home cage to allow mice to self-administer 1 g/kg of alcohol. Following the completion of alcohol or water self-administration, the mice received an i.p. injection of 17 mg/kg CP-154,526 or vehicle (systemic study), or a microinjection of 0.3  $\mu$ g CP-154,526 or aCSF into the DRN (0.25  $\mu$ l over 2 min; intra-DRN study). The 5-HT concentrations were determined with an HPLC system equipped with electrochemical detection (Shimamoto *et al*, 2011). Changes in 5-HT concentrations are expressed as percent change from average baseline samples for each individual. Injection and probe placements for DRN and mPFC were verified microscopically after cresyl violet staining (Figure 1).

## Statistics

The effects of alcohol and CRF-R1 antagonists on composite aggressive and nonaggressive behaviors were compared using within-group comparisons, as all experimental subjects were tested under all treatment conditions. Behaviors were analyzed by ANOVAs with repeated measures, with Drinking (alcohol or water) and Treatment (vehicle and drug doses) as main factors. Frequencies of behaviors were used to compare attack bites and sideways threats, whereas nonaggressive behaviors (walking, rearing, grooming) are shown as a composite score, and compared for their



**Figure 1** Representative photomicrographs and coronal sections from a stereotaxic atlas (Paxinos and Franklin, 2001) showing correct mPFC probe placements (lines (a)) and DRN cannula placements for dialysis mice (circles (b)).

duration. When appropriate, *post hoc* Bonferroni tests were conducted to determine the treatment conditions that differed significantly from vehicle and water conditions. For the *in vivo* microdialysis experiments, one or two-way ANOVAs with repeated measures were carried out for four data points, including one single baseline value computed as the average of three baseline points, and three subsequent data points following drug treatments. For all comparisons,  $p < 0.05$  was considered significant.

## RESULTS

### Systemic Injections of CRF-R1 Antagonists Reduce Both Alcohol-Heightened and Species-Typical Aggression

Alcohol self-administration significantly increased aggressive behaviors, as compared with vehicle trials after mice self-administered water (Figure 2a and b). There was a significant Drinking effect on the numbers of attack bites ( $F_{(1,46)} = 7.910$ ,  $p = 0.012$ ) and sideways threats ( $F_{(1,46)} = 7.776$ ,  $p = 0.012$ ). Systemic injection of CP-154,526 immediately after alcohol or water self-administration significantly decreased aggressive behaviors compared with vehicle conditions in a dose-dependent manner. There was a significant Treatment effect on the number of attack bites ( $F_{(3,46)} = 26.376$ ,  $p < 0.001$ ) and sideways threats ( $F_{(3,46)} = 26.973$ ,  $p < 0.001$ ). A significant reduction in these two aggressive behaviors was detected in mice self-administering alcohol or water when CP-154,526 was administered in doses of 17 or 30 mg/kg ( $p < 0.05$ ). CP-154,526 also significantly reduced duration of motor activity ( $F_{(3,46)} = 3.329$ ,  $p = 0.026$ , Figure 2c), but only at the highest dose (30 mg/kg), and after water drinking.

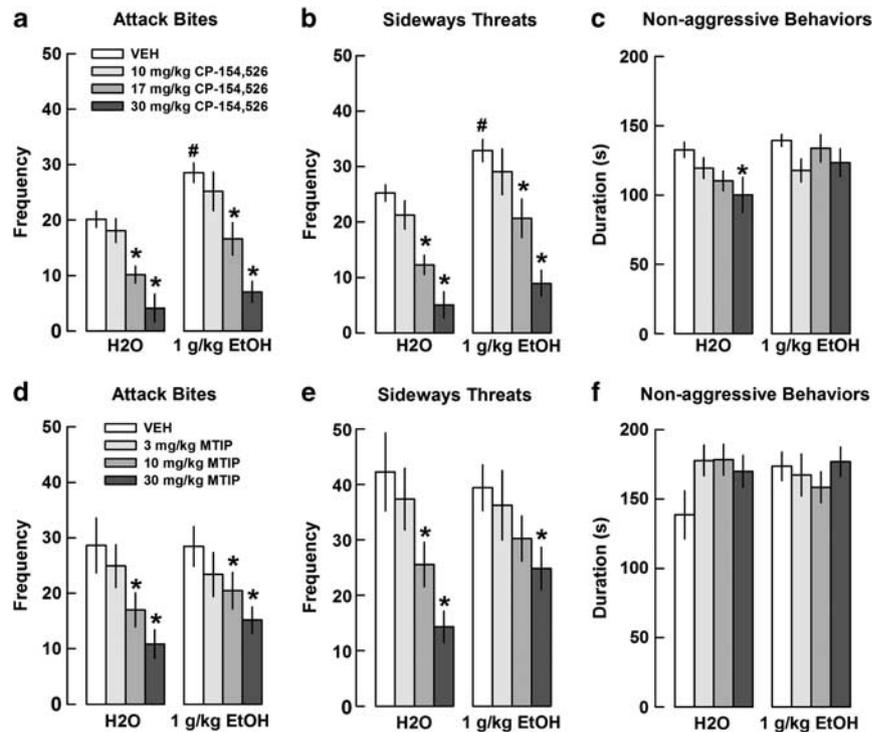
Detailed analysis of nonaggressive behaviors are shown in Supplementary Material (Supplementary Table S1).

MTIP administration in mice before a confrontation significantly reduced aggressive behaviors compared with the vehicle administration (Figure 2d and e), as indicated by a significant Drug effect on the number of attack bites ( $F_{(3,35)} = 9.977$ ,  $p < 0.001$ ) and sideways threats ( $F_{(3,35)} = 9.221$ ,  $p < 0.001$ ). Bonferroni methods showed a significant reduction in these two aggressive behaviors in mice self-administering alcohol when MTIP was administered in a dose of 30 mg/kg ( $p < 0.05$ ). In mice self-administering water, MTIP at both 10 and 30 mg/kg significantly reduced aggressive behaviors ( $p < 0.05$ ). MTIP did not alter motor activity ( $F_{(3,35)} = 0.781$ ,  $p = 0.512$ ). Detailed analysis of nonaggressive behaviors are shown in Supplementary Material (Supplementary Table S1).

### Blockade of CRF-R1 in DRN Selectively Prevents Alcohol-Heightened Aggression

Microinfusion of CP-154,526 into the DRN ( $n = 11$ ) significantly suppressed alcohol-heightened aggression, but had no significant behavioral effects when mice consumed water (Figure 3a; Drinking  $\times$  Treatment interaction in attack bite frequency:  $F_{(2,20)} = 6.02$ ,  $p = 0.009$ ). Both doses of the antagonist, 0.3 and 0.6  $\mu\text{g}$ , reduced the frequency of attack bites when aggressive mice had consumed alcohol before the confrontation ( $p = 0.025$  and  $p = 0.0003$ , respectively). Similar antiaggressive effects were observed for sideways threats (Treatment effect:  $F_{(2,20)} = 10.3567$ ,  $p = 0.0008$ ), particularly when mice were tested after alcohol drinking, as shown in Figure 3b. Importantly, nonaggressive behaviors were not affected by intra-DRN infusion of CP-154,526 in any of the conditions or doses tested (Figure 3c; see Supplementary Table S2 for detailed results on nonaggressive behaviors). These data suggest a selective antiaggressive effect of the CRF-R1 antagonist on alcohol-related aggression. Intra-DRN MTIP ( $n = 13$ ) had effects on aggressive behaviors similar to those of CP-154,526, such that 0.345  $\mu\text{g}$  of MTIP into the DRN reduced the number of attack bites when mice consumed alcohol before the confrontations (Drinking  $\times$  Treatment interaction:  $F_{(1,12)} = 4.74$ ,  $p = 0.0501$ , Figure 3d), with no effects of MTIP when mice were tested during water trials. Similar to CP-154,526, intra-DRN infusion of MTIP did not affect nonaggressive behaviors in any of the conditions tested (Figure 3f; see detailed analysis of nonaggressive behaviors in Supplementary Table S2).

In order to control for possible experience factors that might affect the ability of CRF-R1 antagonism to produce antiaggressive effects (ie, repeated alcohol exposure or chronic water restriction), we next tested the effects of intra-DRN infusion of CP-154,526 in mice with no history of alcohol consumption ( $n = 6$ ). Aggressive mice were tested in confrontations with a male intruder after the infusion of increasing doses of CP-154,526 (0.1, 0.3, and 0.6  $\mu\text{g}$ ) and interspersed vehicle (aCSF) trials. There were no significant effects of CP-154,526 on aggressive or nonaggressive behaviors in mice with no history of alcohol consumption (Table 1). Therefore, the antiaggressive effects of intra-DRN infusion of CRF1 antagonists were selective for conditions



**Figure 2** Systemic CRF-R1 antagonists reduced aggressive behaviors after alcohol and water consumption. Effect of i.p. injections of CP-154,526 ( $n = 18$ , top three panels) or MTIP ( $n = 13$ , bottom three panels) on attack bite frequency (a, d), sideways threats (b, e), and nonaggressive locomotor behavior duration (in s; c, f) after oral self-administration of 1 g/kg ethanol or water. Values are means  $\pm$  SEM; \* $p < 0.05$  compared with water condition, # $p < 0.05$  compared with vehicle test (VEH).

of escalated aggression under the influence of a moderate dose of alcohol.

For this series of intra-DRN experiments, eight mice were excluded because of cannula misplacement (four with cannula tips reaching the aqueduct, two in the median raphe, and two were too lateral). Animals with misplaced cannula failed to present effects of the CRF-R1 antagonist treatment, although statistical analysis was not possible because they were distributed across different drug treatment conditions.

#### Intact 5-HT Activity in the DRN Is Required for the Antiaggressive Effects of CP-154,526

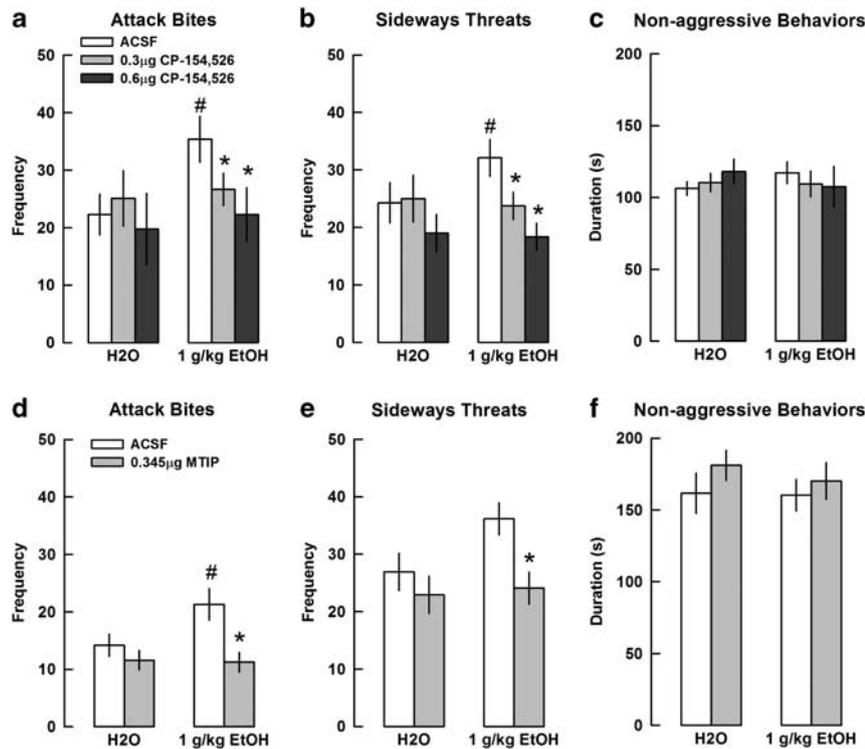
We next tested whether transient inhibition of 5-HT impulse flow, with the coinfusion of a 5-HT<sub>1A</sub> agonist, 0.3  $\mu$ g 8-OH-DPAT, would affect the antiaggressive effects of intra-DRN CP-154,526 on alcohol-related aggression. Intra-DRN infusion of the CRF-R1 antagonist alone was effective in reducing attack bites under the influence of alcohol ( $p = 0.012$  vs vehicle), replicating the preceding experiment (significant Drinking and Treatment interaction:  $F_{(3,27)} = 7.432$ ,  $p = 0.001$ ; Figure 4a). The coinfusion of 0.3  $\mu$ g 8-OH-DPAT into the DRN prevented the emergence of the antiaggressive effects of 0.3  $\mu$ g CP-154,526 on alcohol-heightened aggression ( $p = 0.001$  relative to the alcohol-CP-154,526 alone condition). However, 8-OH-DPAT alone also reduced alcohol-heightened aggression. When the 0.6  $\mu$ g dose of CP-154,526 was used, 0.3  $\mu$ g 8-OH-DPAT could not overcome the antiaggressive effect of CP-154,526 on

alcohol-heightened aggression (Figure 4b). Neither drug alone nor in combination significantly altered aggressive behavior in the absence of alcohol. Nonaggressive behaviors were not affected by any of the treatments (data not shown).

#### 5-HT Release Is Enhanced in the mPFC as a Result of CP-154,526 Action under Alcohol Conditions

Aggression-experienced, alcohol-drinking mice were implanted with the microdialysis guide cannula aimed at the mPFC. During the test, baseline microdialysis samples were collected, and then mice self-administered 1 g/kg alcohol. Immediately after alcohol self-administration, mice received either vehicle (CMC) or the intermediate dose of CP-154,526 (17 mg/kg), i.p., and microdialysis samples continued to be collected for 60 min. There was an overall Group effect on systemic administration of CP-154,526 ( $F_{(1,16)} = 7.568$ ,  $p = 0.032$ ; Figure 5a). The *post hoc* tests showed that the group receiving CP-154,526 after drinking alcohol had higher 5-HT levels only during the first 20 min after treatments when compared with mice receiving vehicle after alcohol consumption. One mouse was excluded because of misplacement of the dialysis probe. The 5-HT levels were not affected by the treatment in this animal.

Following the same experimental design described above, 0.3  $\mu$ g CP-154,526 was infused into the DRN right after completion of the alcohol drinking task. The CRF-R1 antagonist significantly increased prefrontal 5-HT levels after alcohol self-administration compared with baseline ( $p < 0.05$ ). In addition, 5-HT levels were significantly higher



**Figure 3** Intra-DRN CRF-R1 antagonists selectively reduced alcohol-heightened aggression. Effect of intra-DRN microinjections of CP-154,526 ( $n = 11$ , top panels) or MTIP ( $n = 13$ , bottom panels) on frequency of attack bites (a, d) sideways threats (b, e), and nonaggressive locomotor behavior duration (in s; c, f) after oral self-administration of 1 g/kg ethanol or water. Values are means  $\pm$  SEM; # $p < 0.05$  compared with water, \* $p < 0.05$  compared with vehicle.

**Table 1** Behavioral Effects of Intra-DRN Infusion of CP-154,526 in Alcohol-Naive Mice ( $n = 6$ )

		CP-154,526 (in $\mu\text{g}$ )		
aCSF (vehicle)		0.1	0.3	0.6
<i>Aggressive behaviors (frequency)</i>				
Bites	26.3 (4.04)	20.7 (1.71)	26.2 (5.62)	22.0 (4.14)
Threats	36.8 (5.36)	34.8 (3.52)	34.3 (6.92)	32.8 (7.57)
<i>Nonaggressive behaviors (duration in s)</i>				
Walking	147.8 (21.72)	148.3 (11.42)	119.0 (24.61)	139.0 (14.58)
Rearing	24.3 (4.59)	28.2 (6.85)	29.5 (10.89)	26.0 (7.65)
Grooming	10.5 (3.34)	17.7 (3.24)	17.5 (4.81)	20.7 (5.67)

Data shown as averages ( $\pm$  SEM).

in the mice that received CP-154,526 ( $n = 7$ ) compared with the mice that received vehicle treatment ( $n = 5$ ,  $p < 0.05$ ; Figure 5b). In total, four animals were excluded. The 5-HT levels were not affected by the manipulation in these animals.

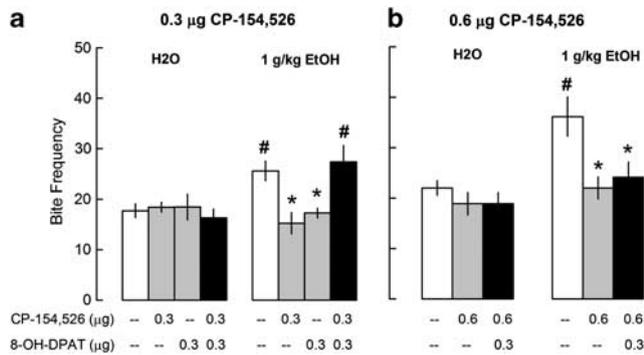
In separate control experiments, we found that 5-HT levels were unaltered in mice that only self-administered 1 g/kg alcohol ( $F_{(5,26)} = 0.378$ ,  $p = 0.859$ ,  $n = 7$ , data not shown) or in mice receiving CP-154,526 alone (17 mg/kg, i.p.), without the combined influence of alcohol ( $F_{(6,11)} = 1.103$ ,  $p = 0.419$ ,  $n = 4$ , data not shown). One animal presented misplaced probe and was not included in the data analysis.

## DISCUSSION

The current set of studies reveals a key role for CRF-R1 in aggression that is escalated by alcohol in male mice via modulation of 5-HT activity in the DRN-mPFC pathway. We found that systemic administration of CRF-R1 antagonists CP-154,526 and MTIP prevented alcohol-heightened aggression and species-typical aggression, whereas infusion of CRF-R1 antagonists into the DRN selectively prevented alcohol-heightened aggression. By transiently inhibiting DRN impulse flow with a low dose of 8-OH-DPAT, we found that intact DRN 5-HT activity is required for a CRF-R1 antagonist to prevent escalated alcohol aggression in a dose-dependent manner. Furthermore, *in vivo* microdialysis studies suggest that increases in 5-HT release in the mPFC are associated with the antiaggressive effects resulting from blockade of CRF-R1 after alcohol drinking.

### Blockade of CRF-R1: Systemic vs DRN Effects on Alcohol-Related Aggression

Selective antiaggressive effects on escalated aggression promoted by alcohol were only observed when CRF-R1 antagonists were locally infused into the DRN, whereas more generalized antiaggressive effects were observed with systemic blockade of CRF-R1. These results suggest that CRF-R1 in areas other than the DRN probably modulate species-typical levels of aggressive behavior, whereas DRN CRF-R1 are particularly important when aggression is performed under the influence of alcohol. In all but one of the several experiments of this study (MTIP systemic



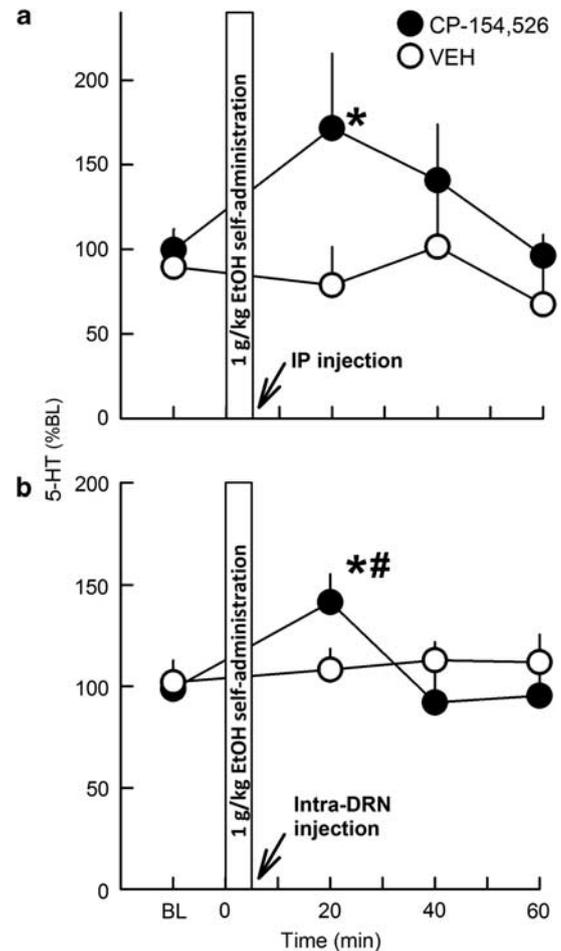
**Figure 4** Intact DRN 5-HT function is required for CRF-R1 reduction of alcohol-heightened aggression. Effect of intra-DRN microinjections of 0.3 µg (a) or 0.6 µg (b) CP-154,526 and 0.3 µg 8-OH-DPAT, 5-HT<sub>1A</sub> agonist, separately and in combination, on attack bite frequency after oral self-administration of 1 g/kg ethanol or water. Values are means ± SEM;  $n = 10$ ; # $p < 0.05$  compared with water. \* $p < 0.05$  compared with vehicle.

experiment), alcohol-heightened aggression was significantly observed in the overall group of animals, despite the known individual differences in this phenomenon (see Miczek *et al*, 2004). Such differences may have contributed to the absence of alcohol-escalated aggression in that particular cohort of mice. However, the antiaggressive effects of the CRF-R1 antagonists were observed in that and all other experiments involving alcohol in this study.

Importantly, CRF-R1 antagonists in the DRN did not affect other nonaggressive motor behaviors. In nonalcohol-drinking mice, intra-DRN blockade of CRF-R1 failed to affect baseline levels of aggressive behavior, supporting the hypothesis that DRN CRF-R1 may be critical only in conditions of escalated aggression (Miczek *et al*, 2013). It remains to be determined whether CRF-R1 blockade in the DRN is also effective in reducing other types of escalated aggression.

Evidence for antiaggressive effects because of antagonism of CRF-R1 have been obtained with another selective, nonpeptidergic CRF-R1 antagonist, SSR125543A, in male Syrian hamsters (Farrokhi *et al*, 2004), as well as with a nonselective antagonist,  $\alpha$ -helical CRF, in stress-induced defensive aggression in rats (Tazi *et al*, 1987). However, CRF-R1 knockout mice exhibited normal levels of intermale aggression compared with wild types (Gammie and Stevenson, 2006). Furthermore, the involvement of CRF-R1 in other types of aggression, such as maternal aggression, seems to be secondary as compared with the role of CRF-R2 (see, eg, D'Anna *et al*, 2005; Gammie *et al*, 2005; Gammie and Stevenson, 2006). The present data show an involvement of CRF-R1 in the modulation of alcohol-heightened aggression in male mice, and point to the DRN as a critical site for these selective antiaggressive effects.

An important role for CRF and its associated peptides, Urocortins 1, 2, and 3, has been proposed in the modulation of several different social behaviors (Hostetler and Ryabinin, 2013). There is evidence that the direct administration of CRF itself may facilitate or induce pro-aggressive and defensive effects after i.c.v. or intra-amygdala infusions in male rats (Elkabir *et al*, 1990; Tazi *et al*, 1987), as well as in rainbow trout (Carpenter *et al*, 2009). However, there is also evidence that the effects of CRF are dose, condition, and



**Figure 5** Systemic and intra-DRN CRF-R1 antagonist increased mPFC 5-HT. Effect of i.p. injection ( $n = 4$ , a) or intra-DRN microinjection ( $n = 7$ , b) of CP-154-526 (filled symbols) or vehicle (open symbols) on 5-HT levels in mPFC after oral self-administration of 1 g/kg ethanol. Shown are 5-HT levels (pg) expressed as percent (%) change from baseline over time (min). Values are means ± SEM; # $p < 0.05$  compared with water, \* $p < 0.05$  compared with vehicle.

species dependent (Elkabir *et al*, 1990; Tazi *et al*, 1987; Mele *et al*, 1987). For example, in isolated male mice, i.c.v. infusion of CRF or sauvagine, a CRF-R2 agonist, significantly reduced aggressive behaviors (Mele *et al*, 1987). Both CRF and Urocortins also have suppressive effects on maternal aggression, mostly dependent on CRF-R2 (D'Anna *et al*, 2005; Gammie *et al*, 2004, 2005).

Although this is the first study on the modulation of alcohol-heightened aggression by CRF-R1 signaling, the involvement of CRF systems in alcohol drinking and alcohol dependence has been extensively explored in animal models (Heilig and Koob, 2007). After repeated and chronic exposure to alcohol, rats show hyperactive extrahypothalamic CRF activity, as indicated by increases in CRF levels and its receptors in the extended amygdala (Merlo Pich *et al*, 1995; Olive *et al*, 2002; Sommer *et al*, 2008). Systemic or intra-amygdaloid blockade of CRF-R1 attenuates increased alcohol seeking and intake in alcohol-dependent animals (Funk *et al*, 2006; Gehlert *et al*, 2007; Sommer *et al*, 2008; Heilig and Koob, 2007) or in high-alcohol-consuming mice (Sparta *et al*, 2008). We also recently observed that CP-154,526 microinjected in the DRN reduced alcohol drinking

in rats and mice that drank excessively, but not in those animals that showed low-level or moderate alcohol drinking (Hwa *et al*, 2013). Sensitized anxiety-like behavior and increased drinking, in the context of withdrawal from alcohol, can also be prevented by CRF-R1 antagonists in the DRN and other sites (Huang *et al*, 2010; Knapp *et al*, 2011). Together, these results and ours indicate that CRF appears to play a role in excessive alcohol-related behaviors, whether it is escalated drinking leading to dependence, or escalated aggression caused by an acute, moderate dose of ethanol.

### CRF-5-HT Interactions in the DRN: a Role in Alcohol-Escalated Aggression

We targeted CRF receptors in the DRN for reducing alcohol-heightened aggression because of the long-standing hypothesis that serotonin activity is critical for the development and expression of pathological aggression (Takahashi *et al*, 2011; Takahashi and Miczek, 2013). Serotonergic pathways, including uptake and storage processes, and somatodendritic and terminal pre- and postsynaptic receptor mechanisms have been investigated for the neural control of aggressive behavior (de Boer and Koolhaas, 2005; Takahashi and Miczek, 2013). Tonic levels of 5-HT are critical determinants in the genesis of impulsive outbursts, while its impulse flow in the DRN can reduce several types of species-specific and maladaptive aggressive behaviors. Phasic changes in 5-HT emerge during aggressive episodes as illustrated by a sudden decrease in PFC 5-HT during a confrontation (Van Erp and Miczek, 2000).

In order to assess the involvement of DRN 5-HT neurons in mediating the effects of CRF-R1 antagonists, we used 8-OH-DPAT as a tool to transiently inhibit DRN 5-HT impulse flow during aggressive behavior. A low dose of 8-OH-DPAT was used so that baseline levels of aggression would be preserved (Takahashi *et al*, 2010b), as higher doses result in behavioral inhibition (Faccidomo *et al*, 2008). As expected, intra-DRN infusion of CP-154,526 abolished alcohol-heightened aggression. In the presence of 8-OH-DPAT, however, the CRF-R1 antagonist failed to prevent alcohol-escalated aggression. These results suggest the requirement of intact 5-HT activity in the DRN for the antiaggressive effects of CP-154,526 to emerge under the influence of alcohol drinking. Intriguingly, the infusion of 8-OH-DPAT alone in the DRN also prevented alcohol-heightened aggression in the absence of the CRF-R1 antagonist. Although 8-OH-DPAT could reverse the lower dose of the CRF-R1 antagonist, it could not reverse the higher dose of the CRF-R1 antagonist, suggesting 5-HT may be more crucial at the lower dose of CP-154,526. The relationship between CRF-R1 and 5-HT<sub>1A</sub> receptors may be complex. The fact that CRF-R1 and 5-HT<sub>1A</sub> receptors can be found in both 5-HT and GABAergic neurons in the DRN (see, eg, Beck *et al*, 2004; Kirby *et al*, 2003) further complicates the understanding of this CRF-5-HT interaction in the DRN, with a putative role for a GABAergic modulation (Takahashi *et al*, 2010b). Thus, the precise mechanism by which 8-OH-DPAT influences alcohol-heightened aggression, and interacts with CRF-R1 receptors, remains to be determined.

Among the forebrain and limbic structures that are innervated by the DRN, the mPFC is likely to be a critical

target for the modulation of aggressive behavior (Olivier, 2004). This pathway is of particular interest in view of dysfunctions found in the mPFC of violent criminals and aggressive psychiatric patients (see, eg, Best *et al*, 2002; Raine *et al*, 1994), as well as in genetically selected highly aggressive rodents (Caramaschi *et al*, 2008). Moreover, *in vivo* microdialysis of male aggressive rats shows a reduction in PFC 5-HT levels during an aggressive confrontation with a male conspecific, suggesting a proximal involvement of 5-HT levels in the display of aggressive behaviors (van Erp and Miczek, 2000). Our *in vivo* microdialysis results suggest that increases in mPFC 5-HT levels are associated with the antiaggressive effects of a CRF-R1 antagonist. Indeed, in alcohol-consuming mice, CP-154,526 administered systemically or directly into the DRN promoted significant increases in 5-HT concentrations in the mPFC. Serotonin concentrations peaked in the first sample after alcohol consumption and drug treatment (within 20 min) in a time frame consistent with the period when antiaggressive effects of CP-154,526 were observed in behavioral studies. Alcohol consumption or CP-154,526 administration alone did not affect mPFC 5-HT levels.

Our findings contrast with those of Takahashi *et al* (2010b) who reported increased 5-HT levels in the mPFC as a result of the GABA<sub>B</sub> agonist baclofen infusion in the DRN in a dose that increases aggression above species-typical levels of aggressive behavior. In our case, consumption of an aggression-heightening dose of alcohol failed to affect 5-HT levels in the mPFC, whereas increased 5-HT was observed when escalated aggression was *prevented* by the CRF-R1 antagonist. Furthermore, mice in our microdialysis experiments were experienced with aggression and alcohol drinking, and both of these factors have been shown to affect 5-HT levels and transmission in the mPFC (see, eg, Caramaschi *et al*, 2008; Faccidomo *et al*, 2008). Mice used in the microdialysis assays in the study of Takahashi *et al* (2010b) were neither experienced aggressors nor alcohol consumers. Together, one could speculate that different 5-HT mechanisms may be recruited under different types of aggression (baclofen *vs* alcohol-heightened aggression), or the prevention thereof. Indeed, a more intricate role for different 5-HT neurons in the modulation of aggression has been proposed (see, eg, de Boer and Koolhaas, 2005; Takahashi *et al*, 2011). Despite presenting a PFC-5-HT mechanism for the antiaggressive effects of CRF-R1 antagonists under alcohol conditions, the present study does not rule out the participation of other brain regions and pathways associated with the emergence of alcohol-heightened aggression.

Electrophysiological and microdialysis studies have consistently shown that i.c.v. or intra-DRN microinjections of CRF, or drugs targeting CRF receptors, exert potent modulatory control over 5-HT neurons (Kirby *et al*, 2000; Lowry *et al*, 2000), modulating 5-HT release in striatum, amygdala, lateral septum, and prefrontal cortex (Amat *et al*, 2004; Forster *et al*, 2008; Lukkes *et al*, 2008; Price *et al*, 1998; Price and Lucki, 2001). Evidence suggests that activation of DRN CRF-R1 predominantly inhibits whereas CRF-R2 activation predominantly increases 5-HT function and output (Valentino and Commons, 2005; Valentino *et al*, 2010). Interpreting our data according to this model, one could speculate that when CRF-R1 antagonists are infused in the DRN after alcohol drinking, blockade of CRF-R1

could cause a shift in CRF actions in favor of CRF-R2 activation. In turn, this shift could contribute to the observed increases in 5-HT concentrations in the mPFC, which is the proposed mechanism for the observed antiaggressive effects of CRF1 antagonists on alcohol-escalated aggression. Ongoing studies are characterizing the involvement of CRF-R2 in the DRN in alcohol-related aggression.

In conclusion, these studies implicate CRF-R1 modulation of the serotonergic DRN-mPFC pathway as a critical mechanism for the antiaggressive effects of CRF-R1 antagonism, particularly in the context of alcohol-heightened aggression in male mice. It would be interesting to determine whether this circuit is also important for other types of escalated aggression such as frustration-heightened aggression or maternal aggression. Understanding the neurobiology of escalated aggression in social animals is a step forward toward pharmacological treatment for intense pathological aggression. This research proposes CRF-R1 antagonists as putative tools for preventing this hazardous behavior, especially under the influence of alcohol. Clinical therapeutic use of CRF-R1 antagonists should be in sight for controlling alcohol-related abuse and aggression, once new and safe bioavailable molecules targeting CRF-R1 are made available.

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